Amendments to the Specification:

Kindly insert the attached sequence listing into the specification:

At page 18, lines 16-34 to page 19, lines 1-3

The fluorescent peptide substrate used in the MMP-12 assay is FAM-Gly-Pro-Leu-Gly-Leu-Phe-Ala-Arg-Lys(TAMRA) **(SEQ ID. No. 1)**, where FAM represents carboxyfluorescein, and TAMRA represents tetramethylrhodamine. MMP12 catalytic domain (residues 106-268) protein was expressed in *E. coli* in the form of insoluble inclusion bodies & stored in concentrated solution under denaturing conditions (8M guanidine hydrochloride). Enzyme was refolded into active form *in situ* by direct dilution into assay reactions. The 51 uL reactions are run in NUNC-brand black, square 384-well plates, each well containing 2 uM substrate, 20 nM enzyme, and 0.001-100 uM inhibitor, in 50 mM HEPES, pH 7.5, 150 mM NaCl, 10 mM CaCl2, 1 uM ZnAc, 0.6 mM CHAPS, and 2 % DMSO. Positive control wells contain no inhibitor. Negative control wells are effected by either pre-dispensing the EDTA quench (see below) or by omitting enzyme. Reactions are incubated at ambient temperature for 120 min, then quenched by the addition of 15uL of 100mM EDTA. Product formation in each well is quantified by measuring flourescense with a Molecular Devices Acquest. The excitation wavelength is set at 485 nM, and the emission wavelength is 530 nM. IC_{50} values were obtained by first calculating the percent inhibition (%I) at each inhibitor concentration (%I = 100*(1-(I-C2)/(C1-C2)), where C1 is the mean of the positive controls, and C2 is the mean of the negative controls), then fitting the %I vs. inhibitor concentration [I] data to: %I=A+((B-A)/(1+((C/[I]^D))), where A is the lower asymptote, B is the upper asymptote, C is the IC50 value, and D is the slope factor. When tested in this assay, compounds of the Examples had IC50s below 100 micromolar.